

## Original Article

# Spinal cord fusion with PEG-GNRs (TexasPEG): Neurophysiological recovery in 24 hours in rats

C-Yoon Kim<sup>1,2,†</sup>, William K. A. Sikkema<sup>4,†</sup>, In-Kyu Hwang<sup>1</sup>, Hanseul Oh<sup>2</sup>, Un Jeng Kim<sup>3</sup>, Bae Hwan Lee<sup>3</sup>, James M. Tour<sup>4,5,6</sup>

<sup>1</sup>Department of Stem Cell Biology, School of Medicine, Konkuk University, Seoul, Korea, <sup>2</sup>Department of Laboratory Animal Medicine, College of Veterinary Medicine, Seoul National University, Seoul, Korea, <sup>3</sup>Department of Physiology, Brain Korea 21 PLUS Project for Medical Science, Yonsei University College of Medicine, Seoul, Korea, <sup>4</sup>Department of Chemistry, <sup>5</sup>The NanoCarbon Center, <sup>6</sup>Department of Material Science and Nanoengineering, Rice University, Houston, Texas, USA

E-mail: C-Yoon Kim - [vivavets@gmail.com](mailto:vivavets@gmail.com); William K. A. Sikkema - [William.Sikkema@rice.edu](mailto:William.Sikkema@rice.edu); In-Kyu Hwang - [weaver0903@naver.com](mailto:weaver0903@naver.com); Hanseul Oh - [seul3198@snu.ac.kr](mailto:seul3198@snu.ac.kr); Un Jeng Kim - [mignon@yuhs.ac](mailto:mignon@yuhs.ac); \*Bae Hwan Lee - [bhlee@yuhs.ac](mailto:bhlee@yuhs.ac); \*James M. Tour - [Tour@Rice.edu](mailto:Tour@Rice.edu)

\*Corresponding author

<sup>†</sup>These authors equally contributed to this work.

Received: 07 August 16 Accepted: 31 August 16 Published: 13 September 16

## Abstract

**Background:** The GEMINI spinal cord fusion protocol has been developed to achieve a successful cephalosomatic anastomosis. Here, for the first time, we report the effects of locally applied water-soluble, conductive PEG(polyethylene glycol)ylated graphene nanoribbons (PEG-GNRs) on neurophysiologic conduction after sharp cervical cord transection in rats. PEG-GNRs were produced by the polymerization of ethylene oxide from anion-edged graphene nanoribbons. These combine the fusogenic potential of PEG with the electrical conducting properties of the graphene nanoribbons.

**Methods:** Laminectomy and transection of cervical spinal cord (C5) was performed on Female Sprague-Dawley (SD) rats. After applying PEG-GNR on the severed part, electrophysiological recovery of the reconstructed cervical spinal cord was confirmed by somatosensory evoked potentials (SSEPs) at 24 h after surgery.

**Results:** While no SSEPs were detected in the control group, PEG-GNR treated group showed fast recovery of SSEPs at 24 h after the surgery.

**Conclusion:** In this preliminary dataset, for the first time, we report the effect of a novel form of PEG with the goal of rapid reconstruction of a sharply severed spinal cord.

**Key Words:** Cephalosomatic anastomosis, electrophysiology, graphene nanoribbons, GEMINI, spinal cord fusion

Video is Available on:  
[www.surgicalneurologyint.com](http://www.surgicalneurologyint.com)

### Access this article online

**Website:**

[www.surgicalneurologyint.com](http://www.surgicalneurologyint.com)

**DOI:**

10.4103/2152-7806.190475

**Quick Response Code:**

## INTRODUCTION

To achieve a successful spinal cord fusion as required during cephalosomatic anastomosis (CSA), an effective technique to assure rapid reinnervation of the body across the divided cervical spinal cord is necessary.<sup>[5]</sup>

In the proposed spinal cord fusion protocol called GEMINI, reapposition of two sharply severed cords

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

**For reprints contact:** [reprints@medknow.com](mailto:reprints@medknow.com)

**How to cite this article:** Kim CY, Sikkema WK, Hwang IK, Oh H, Kim UJ, Lee BH, et al. Spinal cord fusion with PEG-GNRs (TexasPEG): Neurophysiological recovery in 24 hours in rats. *Surg Neurol Int* 2016;7:S632-6.  
[http://surgicalneurologyint.com/Spinal-cord-fusion-with-PEG-GNRs-\(TexasPEG\):-Neurophysiological-recovery-in-24-hours-in-rats/](http://surgicalneurologyint.com/Spinal-cord-fusion-with-PEG-GNRs-(TexasPEG):-Neurophysiological-recovery-in-24-hours-in-rats/)

brings in contact the gray matter cores in which the cortico-truncoreticulo-propriospinal (CTRPS) pathway courses; this cellular core re-establishes contact by regrowth of the severed connections among propriospinal cells and by acute “fusion” of the neural membranes that were transected acutely. This regrowth has been confirmed histologically in animal models of spinal cord sections by Freeman (see in reference 6), who demonstrated a huge amount of fibers regrowing across the stumps interface, and more recently by other groups (see in 3-4,6). The CTRPS pathway – rather than the myelinated fibers of the descending and ascending tracts – is truly responsible for motor and coarse sensory transmission and is the linchpin of the GEMINI spinal cord fusion protocol (for an in-depth review of the key role of the evolutionarily conserved CTRPS in sensorimotor integration, refer to references 3–6).

The GEMINI fusion process achieves accelerated fiber fusion by way of substances called fusogens.<sup>[5,11]</sup> This fusion of transected axons is made possible by exploiting fusogens, such as polyethylene glycol (PEG), i.e., substances that restore the integrity of acutely transected nerve fibers both in peripheral nerves and sharply transected spinal cords.<sup>[5,6,11]</sup> This is accompanied by a resealing (sealant effect of PEG) of the membranes of neurons injured by the nanoblade transection (akin to a neuroprotective effect) in the CTRPS spinal core. As mentioned, neuronal survival at the interface of the two apposed stumps is followed by resprouting and restoration of mechanical integrity. This specific process can be accelerated by electrical stimulation.<sup>[4,6]</sup>

Despite the properties of PEG having been discovered in 1986, very little work has been published over the subsequent 30 years.<sup>[5,11]</sup> Acute transections in man being rare, no group pursued a clinical application of this technology. A head-body transplant and the need for an accelerated spinal cord fusion protocol resurrected these older studies.

Fusogens exert their beneficial effect maximally when applied locally to the point of transection rather than when administered parenterally.<sup>[11]</sup> Membrane fusion and attendant mixing of the cytoplasm of fused cells occurs when adjacent membranes touch in the presence of PEG or a similar compound. Acute dehydration of the fusing plasmalemmas permits glycol/protein/lipid structures to resolve into each other at the outer membrane leaflet first and the inner membrane leaflet subsequently. In other words, dehydration of the membrane facilitates the hydrophobic core of the lamellae to become continuous; rehydration after PEG exposure permits the polar forces associated with the water phase to help reorganize the structure of transmembrane elements. PEG is dislodged once the membrane is sealed. This reorganization of

cellular water is believed to result from the strongly hydrophilic structure of PEG.<sup>[5]</sup> This fusion effect was first demonstrated in single axons, then in peripheral nerves.<sup>[5,11]</sup>

In light of these data, we asked whether results could be improved by adding conductive, high-aspect ratio graphene nanoribbons (GNR) to the fusogen solution. In a three-dimensional tissue setting, across a gap in the spinal cord, we expected the GNRs to first act as an electrical conduit and then act as an electrically active scaffold upon which the neurons will grow, directing their processes in the proper direction across the gap. We expect this result primarily because GNRs have been patterned on two-dimensional surfaces cause growing and differentiating neurons to take on the same pattern.<sup>[1,9]</sup> Only a very small concentration of PEG-GNRs in the fusogen was required because at <1 % m/v of high aspect ratio rod-like carbon nanostructures in polymers, electrical percolation (i.e., the concentration at which conductive particles make the overall, usually insulating, solution conductive) can be achieved.<sup>[2]</sup> In other words, in order to improve and accelerate the recovery of function, we tested PEG enhanced by these electrical conducting nanoribbons. PEG-GNRs would achieve both membrane fusion, facilitate initial electrical conduction, and then act as a scaffold for sprouting fibers.

We have recently shown how a sharp transection aided by PEG alone can restore at least partial motor function in a rat and mice<sup>[7,11]</sup> and neurophysiologic transmission<sup>[11]</sup> compared with the control groups of animals.

In this paper, we report for the first time on the effects of a locally applied water-soluble, PEGylated conductive GNR solution on neurophysiologic conduction after sharp cervical cord transection in rats.

## MATERIALS AND METHODS

### Polyethylene glycol–graphene nanoribbon (TexasPEG)

Multi-walled carbon nanotubes (MWCNTs) were obtained from EMD Merck (produced by Mitsui and Co., lot no. 2699-64E) and were used as received. Tetrahydrofuran (THF) was dried over solid KOH for several days, degassed, and freshly distilled from sodium/benzophenone under a N<sub>2</sub> atmosphere. All chemicals were purchased from Sigma-Aldrich unless otherwise specified. Thermogravimetric analysis (TGA) measurements were performed on a TA instruments Q-600 Simultaneous TGA/DSC. The temperature was ramped at 10°C/min until 900°C under Ar. For transmission electron microscopy (TEM) analysis, the PEG-GNRs were dispersed in water and drop cast onto a lacey carbon grid. For scanning electron microscope (SEM) analysis, the PEG-GNRs were dispersed in *o*-dichlorobenzene,

briefly sonicated in a bath sonicator, and deposited on a smooth metal disk, from which the solvent was evaporated on a heat plate at  $<100^{\circ}\text{C}$ . The sample was imaged by an FEI Quanta 400 ESEM FEG instrument. One gram of Mitsui MWCNTs was added to a 1 L oven-dried, nitrogen-purged, Schlenk flask; 500 mL of THF was added. Eutectic NaK (2.5 mL) 1:3.3 by mass (1:1.9 by mol) was added under  $\text{N}_2$ . The reaction mixture was stirred at room temperature for 3 days, until very few liquid droplets of NaK remained. The reaction was cooled in a dry ice/acetone bath to  $-78^{\circ}\text{C}$ , and 30 g (0.7 mol) of gaseous ethylene oxide was added from a lecture bottle over 90 min. The mixture was slowly brought to room temperature and stirred for 3 days. A mixture of NaH (20 mmol, 0.53 g) and propargyl bromide (20 mmol, 2.4 g) suspended/dissolved in dry toluene was added to terminate the ethylene oxide polymerization. The reaction was quenched by the addition of 20 L of water, and the dark grey precipitate was collected via filtration on a  $0.22\ \mu\text{m}$  polyethersulfone (PES) membrane. The dark grey precipitate was filtered through a polytetrafluoroethylene (PTFE) membrane ( $0.45\ \mu\text{m}$ ), followed by crossflow filtration with a 50 kDa MWCO PES filter to remove unbound polymer. The PEG-GNRs final product (1.3 g) was collected on a PTFE membrane ( $0.45\ \mu\text{m}$ ), washed with DI water ( $3 \times 100\ \text{mL}$ ), ethanol ( $3 \times 100\ \text{mL}$ ), DI water ( $3 \times 100\ \text{mL}$ ), and dried under high vacuum overnight. The propargyl units were added to some of the termini for future peptide additions if desired. Prior to use, the PEG-GNRs were dispersed in PEG 600 (0.5 wt% by GNR concentration). The mixture was tightly sealed in a 50 mL conical vial and was sterilized by  $120^{\circ}\text{C}$  pressurized steam for 30 min.<sup>[8,10]</sup>

### Surgery

The experiment was carried out in accordance with animal ethics guidelines and was approved by the Institutional Animal Care and Use Committee of Konkuk University (Seoul, South Korea). Female Sprague-Dawley rats (250–280 g, Young Bio, Gyeonggi-do, Korea) were anesthetized using zoletil and xylazine (3:1 ratio, 1 mL/kg). The muscles overlying the cervical vertebral column were reflected exposing C4-6; a C5 laminectomy was performed and the dura mater was split open longitudinally. After gently raising the cervical cord with a hook, full severance was performed with surgical sharp blades #11.

The experimental group ( $n = 5$ ) was treated with the PEG-GNR solution (0.5 mL per animal) directly applied at the level of transection of the cervical cord. The control group ( $n = 5$ ) was treated with the same volume of phosphate buffered saline. The muscle and fascia were sutured and the skin closed. Dextrose 5% solution (20 mL/kg) was administered daily via intraperitoneal injection. The two stumps of the spinal cord of the rats were kept in mechanical proximity by simple hyperextension of the head.

### Electrophysiology

One day after the surgery, the animals were reanesthetized with urethane (1.25 g/kg, i.p.). Each animal was placed on a stereotaxic device (Narishige Scientific Instrument Laboratory, Tokyo, Japan) and artificially ventilated using a small animal respirator (Model 683, Rodent Ventilator, Harvard, Holliston, MA, USA). Somatosensory evoked potentials (SSEPs) were recorded to measure the conduction recovery of the sensory system. A special electrode (NE-120, Rhodes Medical Instruments, Inc., Woodland Hills, CA, USA) was used for SSEP recording. For the SSEP recording, the recording electrode was placed in the sensorimotor cortex (bregma:  $-2\ \text{mm}$ , lateral:  $2\ \text{mm}$ ; recording side). A bipolar platinum wire electrode placed in the contralateral sciatic nerve (stimulating side) was used as a stimulating electrode.

A single square pulse (0.1 ms duration) of electrical stimulus was delivered by a stimulus isolator (A365D, World Precision Instruments, Inc., New Haven, CT, USA) that was driven by a pulse generator (Pulsemaster A300, World Precision Instruments). The analog signals of the evoked potentials were amplified ( $\times 10000$ ), filtered (Band-Pass 300–1000 Hz), and fed to an IBM-compatible PC through an AD/DA converter (CED 1401, Cambridge, UK) to be averaged out using Spike 2 software. SSEP consisted of an average of 100–300 single sweep epochs. The effect of the stimulation intensity on SSEPs was analyzed in the wave forms by latencies and amplitudes.

### Behavioral assessment

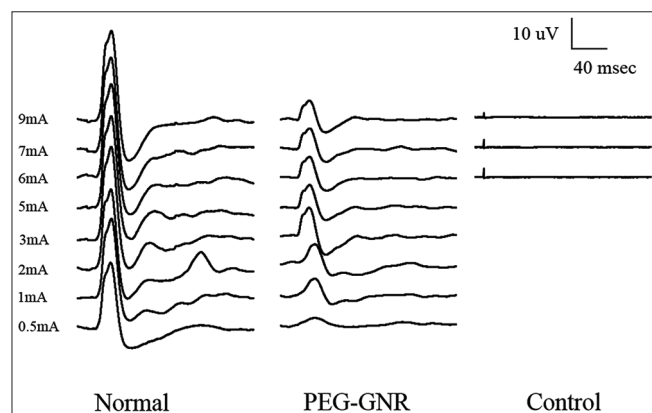
Four of the PEG-GNR treated rats died accidentally (drowning during a storm that filled the underground lab) subsequently to the SSEP study. The surviving rat is reported here versus controls treated with saline. The modified Basso, Beattie, and Bresnahan (mBBB) 22-point (0–21) scoring was employed. A score of 21 indicates unimpaired locomotion as observed in uninjured animals, whereas a score of 0 indicates complete absence of voluntary movement. Assessment occurred daily for 2 weeks.

## RESULTS

SSEPs were used to evaluate the functional integrity of ascending sensory pathways following surgery and topical treatment. SSEPs are a quantitative way to assess the conduction of somatosensory pathways following cervical cord transection. SSEPs were measured from normal rats (without transection) to acquire baseline recordings for comparison. Figure 1 shows the representative wave forms of the SSEPs evoked by stimuli of various intensities. With a stimulus of 6 mA, the amplitude and latency parameters were classified as initial, N1- and P1-peak, and represented the amount of signal transduction

**Table 1: In vivo electrophysiology analysis**

Group	Latency (ms)			Amplitude ( $\mu$ V)		
	Initial	N1	P1	Initial	N1	P1
Normal	9.70 $\pm$ 0.98	21.33 $\pm$ 3.39	39.87 $\pm$ 4.77	0.62 $\pm$ 0.36	58.28 $\pm$ 13.68	105.84 $\pm$ 32.72
PEG-GNR	9.70 $\pm$ 0.75	26.43 $\pm$ 2.22	49.33 $\pm$ 4.62	0.21 $\pm$ 0.05	6.03 $\pm$ 2.63	8.98 $\pm$ 3.75
Control	-	-	-	-	-	-



**Figure 1: Somatosensory evoked potentials (SSEPs) waveforms by 6-mA stimulation in the experimental group 24 hours after transection. No SSEPs were detected in the control group. The PEG-GNR-treated group exhibited SSEP recovery, although not yet at a normal level**

through the site of severance and the speed at which the signal travels, respectively [Table 1]. The treated animals showed a clear recovery of SSEP at 24 hours, whereas controls showed no recovery whatsoever.

In the behavioral assessment portion of this study, recovery in the surviving rat was steady. Twenty-four hours after the surgery, weak tremors of the forelimbs were apparent. Two days later, slight voluntary movements of all 4 paws were occasionally present (mBBB score of fore limbs and hind limbs: 4, 4 respectively). At 1 week [Video], active movements were clearly evident, but balance was not possible (mBBB score of fore limbs and hind limbs: 8, 9 respectively). Two weeks later [Video], the rat could walk without losing balance, stand up on his hind limbs and use his forelimbs to feed himself with pellets (mBBB score of forelimbs and hindlimbs, 19 and 20, respectively). No recovery was observed in controls.

## DISCUSSION

In this preliminary report, we show how the conductive GNR additive to PEG<sup>[8,10]</sup> appears to afford a quick recovery of neurophysiologic sensory transmission in the spinal cord versus none in controls. The behavioral recovery in the surviving rat was remarkable, versus none in the controls. This rat was not submitted to histological analysis because this preliminary study was a purely electrophysiological and behavioral study to test the potential of this enhanced PEG.

As made clear elsewhere,<sup>[4]</sup> the faster the sensorimotor recovery in the injured patient, the fewer complications from prolonged immobilization. Animal studies have shown that restitution of membrane continuity takes many minutes of absolute immobility of the axon segments in the presence of PEG and even more time for the repair to become permanent.<sup>[3-5]</sup> This immobility can be achieved by stabilizing the point of transection with a negative pressure microconnector within which PEG is circulated;<sup>[6]</sup> this device allows immediate delivery of the selected fusogen combined with a stable mechanical apposition for extended time until mechanical stability is achieved.

In our case, in order to mimic the gentle apposition afforded by this connector, PEG-GNRs were infused into the gap left by the sharp transection at C5 in which the two stumps of the spinal cord of rats were kept in mechanical proximity by simple hyperextension of the head.

In this preliminary study, PEG certainly fused a quota of transected axons, which is responsible for the recovery of SSEPs. However, given the much quicker neurophysiological recovery as compared with a similar PEG-only study that assessed SSEP after full dorsal transection,<sup>[11]</sup> it is likely that the nanoribbons supported at least part of this ultra-fast recovery.

Evidence suggests that molecular weights of PEG of <1000 Daltons may be toxic in humans.<sup>[5]</sup> PEG-GNRs have no apparent potential to trigger acute or chronic toxicity. No side effect was observed in this preliminary study.

## CONCLUSION

In conclusion, we report for the first time the effect of a novel form of PEG with the goal of rapid reconstruction of a sharply severed spinal cord. Ongoing studies will clarify its full potential.

## Acknowledgement

The authors thank Prof. Canavero for his invaluable assistance in analyzing the data and coordinating the centers involved in the study.

## Financial support and sponsorship

The research was partially supported by the National Research Foundation of Korea (NRF) Grant (2015R1C1A1A02037047).

## Conflicts of interest

There are no conflicts of interest.

## REFERENCES

1. Akhavan O, Ghaderi E. Differentiation of human neural stem cells into neural networks on graphene nanogrids. *J Mater Chem B* 2013;1:6291-301.
2. Bauhofer W, Kovacs JZ. A review and analysis of electrical percolation in carbon nanotube polymer composites. *Compos Sci Technol* 2009;69:1486-98.
3. Canavero S. Commentary. *Surg Neurol Int* 2015;6:103.
4. Canavero S. The "Gemini" spinal cord fusion protocol: Reloaded. *Surg Neurol Int* 2015;6:18.
5. Canavero S. HEAVEN: The head anastomosis venture Project outline for the first human head transplantation with spinal linkage (GEMINI). *Surg Neurol Int* 2013;4(Suppl 1):S335-42.
6. Canavero S, Ren X, Kim CY, Rosati E. Neurologic foundations of spinal cord fusion (GEMINI). *Surgery* 2016;160:111-9.
7. Kim CY. PEG-assisted reconstruction of the cervical spinal cord in rats: Effects on motor conduction at 1 h. *Spinal Cord* 2016 [Epub ahead of print].
8. Kim ND, Metzger A, Hejazi V, Li Y, Kovalchuk A, Lee SK, et al. Microwave Heating of Functionalized Graphene Nanoribbons in Thermoset Polymers for Wellbore Reinforcement. *ACS App Mater Interfaces* 2016;8:12985-91.
9. Lalwani G, Patel SC, Sitharaman B. Two- and Three-Dimensional All-Carbon Nanomaterial Assemblies for Tissue Engineering and Regenerative Medicine. *Ann Biomed Eng* 2016;44:2020-35.
10. Lu W, Ruan G, Genorio B, Zhu Y, Novosel B, Peng Z, et al. Functionalized graphene nanoribbons via anionic polymerization initiated by alkali metal-intercalated carbon nanotubes. *ACS Nano* 2013;7:2669-75.
11. Ye Y, Kim CY, Miao Q, Ren X. Fusogen-assisted rapid reconstitution of anatomophysiological continuity of the transected spinal cord. *Surgery* 2016;160:20-5.